



Frequency of alterations in the *MEFV* gene and clinical signs in familial Mediterranean fever in Central Anatolia, Turkey

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ABSTRACT. Familial Mediterranean fever is a recessive autoinflammatory disease that is frequent in Armenians, Jews, Arabs, and Turks. The *MEFV* gene is responsible for this disease. We looked for *MEFV* gene variations (polymorphism and mutations) in a population that resides in Central Anatolia, Turkey. DNA was extracted from peripheral blood leukocytes of 802 familial Mediterranean fever patients. The DNA sequence data were examined for approximately 150 different mutations and polymorphisms, including single nucleotide polymorphisms in different exons of the *MEFV* gene. The male:female ratio of these patients was 1.44:1. Mutations were detected in 48.1% of the patients; 7.5% were homozygous, 11.1% were compound heterozygous and 31.5% had only one identifiable mutant allele. No mutations were detected in 51.9% of the patients. The main clinical characteristics of the patients were: abdominal pain in 20.6%, arthritis in 22.9%

and amyloidosis in 4.6%. Sixty-six percent of patients had a family history of familial Mediterranean fever; 19.4% of the patients were found to have parental consanguinity. We conclude that the genetics of familial Mediterranean fever is more complex than has previously been reported; heterozygous patients presenting a severe phenotype should be further analyzed for less common secondary MEFV mutations, using gene sequencing.

Key words: *MEFV* gene; DNA sequencing; Genetics; Mutation; Familial Mediterranean fever

INTRODUCTION

Familial Mediterranean fever (FMF: OMIM #249100) is an autosomal recessive disorder characterized by recurrent attacks of pleuritis, febrile peritonitis and synovitis. It affects mainly Turks, Arabs, Armenians, and non-Ashkenazi Jews (Medlej-Hashim et al., 2010). A familial Mediterranean fever (*MEFV*) gene has been identified on the short arm of chromosome 16 and several mutations in this gene have been demonstrated in FMF patients (Consortium TFF, 1997; Consortium TIF, 1997). The *MEFV* gene consists of 10 exons and encodes a 781-amino acid protein called pyrin, which is expressed in polymorphonuclear cells, cytokine-activated monocytes, dendritic cells, and synovial fibroblasts (Centola et al., 2000; Diaz et al., 2004). Because of the restricted expression of pyrin in innate immune cells, the major role of pyrin appears to be the regulation of inflammation (Kastner, 2005). Mutations interfere with the pyrin domain, allowing an uninterrupted inflammatory cascade. To date more than 150 mutation gene alterations located in the *MEFV* gene have been identified (Öztürk et al., 2008). Five founding mutations M694V, V726A, M680I, M694I (in exon 10), and E148Q (in exon 2) are the most common MEFV mutations (Sarrauste De Menthère C, 2003; Milhavet et al., 2008). Several reports have shown that the M964V mutation is associated with severe disease featuring early onset, high frequency of attacks, the need for the high doses of colchicine and high frequency of amyloidosis in untreated patients (Saatçi et al., 1997; Mattitt et al., 2006; Düşünsel et al., 2008; Paşa et al., 2008; Jarjour, 2010). The syndrome also features high levels of erythrocyte sedimentation rate, C-reactive protein and serum amyloid A, but case-specific consensus on phenotype and genotype in FMF patients has not been reached (Aldea et al., 2004). The carrier frequency of MEFV mutations is quite high in the 4 classically affected populations, ranging from 37 to 39% in Armenians and Iraqi Jews to 20% in Turks, North African and Ashkenazi Jews and Arabs (Pras and Kastner, 1997; Shinar et al., 2000; Schouten et al., 2002, van Gijn et al., 2008).

All molecular genetic methods for detecting mutations are based on differences in the DNA sequence. Logically, DNA sequencing would appear to be the best approach to determining the mutations. DNA sequencing was originally performed by using radioactive labels for the detection of the reaction products, an approach that is unsuitable for clinical use. Current DNA sequencing protocols use fluorescent nucleotides to label the DNA. The sequence is then read with an automated instrument. DNA sequencing generally begins with polymerase chain reaction (PCR) amplification of DNA regions

of interest, followed by sequencing reactions with the PCR products. Instruments that perform automated analysis of DNA sequencing gels are based on real-time detection of fluorescence-labeled sequencing reaction products (Olive and Bean, 1999). In this study, we examined the entire MEFV coding sequence of symptomatic FMF patients to look for mutations potentially missed by currently used screening methods. This study is the first to perform DNA sequence analysis on the whole MEFV coding sequence in all subjects.

PATIENTS AND METHODS

Patients

A total of 802 patients came to our laboratory because of FMF symptoms - especially abdominal, renal and arthralgia symptoms - between June 2008 and December 2010. They were referred from various clinics with specialties in rheumatology, urology, nephrology, and gastroenterology, among others. The mean age of the patients was 30.7 years. Written consent was obtained from all participants, and the review board at our institution approved the study.

Diagnostic techniques

We drew 5 mL blood from each patient into tubes containing ethylenediamine-tetraacetic acid. DNA was extracted using a commercial kit (QIAamp DNA mini kit; Qiagen, Hilden, Germany). Sequencing was performed using a 48-well capillary machine (MegaBACE, Amersham) for the analysis of the whole MEFV coding sequence. All MEFV gene exons were screened for causative mutations via direct DNA sequencing so as not to miss any mutations. We have applied this technique to approximately 150 different mutations and polymorphisms including single-nucleotide polymorphisms in different exons of the MEFV gene. The sequencing data were analyzed using Sequencher 4.6 (Gene Codes, Ann Arbor, MI, USA). All nucleotide alterations were distinguished using this method.

RESULTS

The study included 802 patients with a male:female ratio of 1.44:1 (473 females, 329 males; mean age, 25.0 ± 11.25 years; range, 5-70 years). Mutations were detected in 386 (48.1%) patients; 53 (7.5%) were homozygous, 87 (11.1%) were compound heterozygous, and 246 (31.5%) had only one identifiable mutant allele. No mutations were detected in 416 (51.9%) patients (Table 1). The age of the male patients ranged from 5 to 75 years. The mean age was 31 years. The age of the female patients ranged from 3 to 72 years. The mean age was 29 years. The mean ages of the patients with and without FMF mutations regardless of gender were 30 and 31 years, respectively. The mean age of patients with MEFV gene mutations was 30 years.

The main clinical characteristics of the patients were as follows: abdominal pain in 20.6%, arthritis in 22.9%, and amyloidosis in 4.6%. Sixty-six percent of patients had a positive family history of FMF. The presence of parental consanguinity was also deter-

mined for all patients, and it occurred in 19.4% of patients. *MEFV* mutations were studied using a whole *MEFV* gene sequencing method. No mutations were detected in 416 patients (51.9%; Table 2). Results in 802 FMF patients revealed that M694V was the most frequent mutation (28%), followed by V726A (11.7%), E148Q (9.1%), and M680I(G/C) (3.9%) in the Central Anatolian population (Figure 1). The rarest mutations were heterozygous forms of F479L (in exon 5), M694I (in exon 10), E225D, E230K, T267I (all three in exon 2), I591T (in exon 9), and combined heterozygous forms of E148Q/V726A, E148Q/M694I, E167D/M694V, E167D/F479L (E167D is in exon 2), M694V/K695R, M694V/F479L, M694V/M694I, M694I/V726A, and M694V/P369S. The frequency of the rare mutations was 0.03% (only one patient had all of these mutations). Most of the rare mutations were heterozygous. Four common mutations (M694V, E148Q, M680I(G/C), and V726A) were found in 52.7% of patients (Figure 2). The study population had a high rate of carriers.

Table 1. Clinical symptoms and results of mutation analysis of the patients with different *MEFV* gene mutations.

Mutation		Number of patients with different FMF symptoms (N)		
Allele 1	Allele 2	Abdominal pain	Arthritis	Amyloidosis
M694V	V726A	8	8	6
M694V	M694V	13	13	7
V726A	V726A	2	-	-
E148Q	E148Q	4	1	-
A744S	-	7	8	-
V726A	-	23	15	7
M694V	-	49	54	6
M680I(G/C)	-	8	6	1
M680I(G/C)	M680I(G/C)	3	5	1
P369S	-	5	3	-
E148Q	-	11	22	2
E148Q	M694V	3	9	1
M680I(G/C)	M694V	5	5	1
K695R	-	5	3	-
M680I(G/C)	V726A	6	3	3
M694V	R761H	3	-	-
E148Q	P369S	1	7	-
T267I	-	1	-	-
M694V	F479L	1	-	-
E225D	-	1	-	-
I259V	-	1	-	-
V726A	R761H	1	1	-
M694I	-	1	-	-
R761H	-	1	4	-
A744S	A744S	1	3	-
F479L	-	1	-	-
P369S	M694V	-	1	-
E167D	M694V	-	1	-
M694V	K695R	-	1	-
M680I(G/C)	R761H	-	3	-
E148Q	M694I	-	1	-
E148Q	V726A	-	1	-
E148Q	M680I(G/C)	-	1	2
I591T	-	-	1	-
E230K	-	-	1	-
E167D	F479L	-	2	-
M694I	M694V	-	1	-
Total		165	184	37

Table 2. Genotypes and frequency of detected mutations in Turkish population.

Mutation type	Genotype	Patients	
		N	%
Heterozygous	M694V	108	28.0
	E148Q	35	9.1
	M680I(G/C)	15	3.9
	V726A	45	11.7
	A744S	15	3.9
	P369S	8	2.1
	R761H	5	1.3
	F479L	1	0.3
	M694I	1	0.3
	K695R	8	2.1
	E225D	1	0.3
	E230K	1	0.3
	I259V	1	0.3
	I591T	1	0.3
	T267I	1	0.3
	Total	246	63.8
	Compound heterozygous	E148Q/M680I(G/C)	3
E148Q/V726A		1	0.3
E148Q/M694V		13	3.4
E148Q/P369S		8	2.1
E148Q/M694I		1	0.3
E167D/M694V		1	0.3
E167D/F479L		2	0.5
M680I(G/C)/V726A		12	3.1
M680I(G/C)/R761H		3	0.8
M680I(G/C)/M694V		11	2.8
M694V/R761H		3	0.8
M694V/K695R		1	0.3
M694V/F479L		1	0.3
M694V/M694I		1	0.3
M694I/V726A		1	0.3
M694V/P369S		1	0.3
M694V/V726A		22	5.7
V726A/R761H		2	0.5
Total		87	22.5
Homozygous		A744S	4
	E148Q	5	1.3
	M680I(G/C)	9	2.3
	M694V	33	8.5
	V726A	2	0.5
	Total	53	13.7
	Patients with MEFV mutations	386	48.1
Patients without MEFV mutations	416	51.9	
Total	802	100.0	

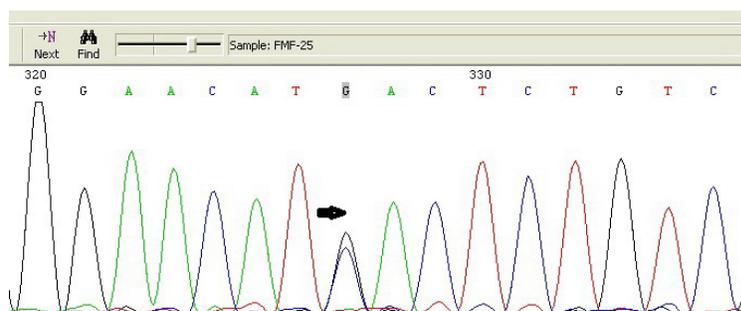


Figure 1. Chromatogram of a patient with M680I(G/C) heterozygosity (arrow).

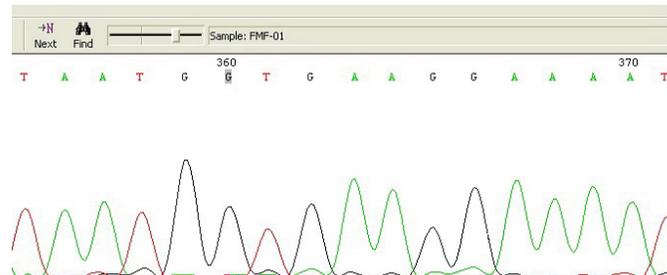


Figure 2. Chromatogram of a patient with M694V homozygosity.

In addition to 7 synonymous polymorphisms in exon 2 (D102D, G138G, and A165A), exon 3 (R314R), and exon 5 (E474E, Q476Q, and D510D), we found a T267I mutation in one heterozygous patient, E225D in one heterozygous patient, E230K in one heterozygous patient, I259V in one heterozygous patient, I591T in one heterozygous patient, F479L in one heterozygous patient, and M694I in one heterozygous patient. These additional mutations were found in different patients and these patients were severe cases of FMF (Table 3). These mutations are nucleotide variations located in exons 2, 5, 9, and 10 that are not explored with denaturing gradient gel electrophoresis or reverse dot methods.

DISCUSSION

The identification of the *MEFV* gene and its various mutations provides a rational basis for medical and genetic counseling for the clinical treatment of FMF patients and their families. The prevalence of this disease in Turkey is approximately 0.1%, but it is estimated that the prevalence may be higher because many patients remain undiagnosed, particularly if they have mild forms of the disease. Furthermore, in Turkey, especially central Anatolia, in which consanguinity is widespread, the incidence of FMF may be higher than that observed. Ankara is situated in the center of Anatolia and receives migration from Central Anatolia, East Anatolia, and the Black Sea region. It is also a referral center for patients from those regions, and our laboratory is one of the main referral institutions. Therefore, the results of our study are representative of the populations living in those regions of Turkey. Most studies of FMF have reported that the disease affects both genders in a similar ratio (Saatçi et al., 1997; Milhavet et al., 2008). *MEFV* gene mutations were similar in male and female FMF patients in our study. Likewise, clinical characteristics were generally comparable between males and females, although some features, such as abdominal pain or arthritis, occasionally differ.

MEFV gene mutations vary according to population characteristics such as familial history, parental consanguinity, migration, population type, and the presence of heterozygous carriers. Both the types and the frequency of mutation in such populations should be examined to protect the health of future generations (Marek-Yagel et al., 2009).

In addition to clinical criteria, molecular studies for detecting disease-causing mutations are needed to confirm the diagnosis of FMF. Sequence analysis is quite precious in FMF diagnoses. We suggest this reliable and safe method for diagnosis in large FMF patient groups. The analysis of the whole *MEFV* gene has shown that M694V is the most frequent mutation in the population from Central Anatolia. Analysis of all mutations in the *MEFV* gene confirmed

Table 3. MEFV genotypes and allele frequencies among 802 FMF Turkish patients.

A. Genotypes		
Genotype		Number of patients
Allele 1	Allele 2	
E148Q	P369S	8
E148Q	M680I(G/C)	3
E148Q	M694V	13
E148Q	M694I	1
E148Q	V726A	1
E167D	F479L	2
E167D	M694V	1
M680I(G/C)	M694V	11
M680I(G/C)	R761H	3
M680I(G/C)	V726A	12
M694V	F479L	1
M694V	K695R	1
M694V	M694I	1
M694V	P369S	1
M694V	R761H	3
M694V	V726A	22
M694I	V726A	1
V726A	R761H	2
M694V	wt	108
E148Q	wt	35
M680I(G/C)	wt	15
V726A	wt	45
A744S	wt	15
P369S	wt	8
R761H	wt	5
F479L	wt	1
M694I	wt	1
K695R	wt	8
E225D	wt	1
E230K	wt	1
I259V	wt	1
I591T	wt	1
T267I	wt	1
A744S	A744S	4
E148Q	E148Q	5
M680I(G/C)	M680I(G/C)	9
M694V	M694V	33
V726A	V726A	2
wt	wt	416
Total		802
B. Allele frequencies		
Mutation	No. of alleles	Frequency
E148Q	71	0.044
E167D	3	0.002
M680I(G/C)	62	0.040
M694V	228	0.142
M694I	3	0.002
V726A	87	0.054
R761H	13	0.008
A744S	23	0.014
K695R	9	0.006
F479L	4	0.003
P369S	17	0.010
E225D	1	0.0006
E230K	1	0.0006
I259V	1	0.0006
I591T	1	0.0006
T267I	1	0.0006
Total mutant	525	0.327
Wild type (wt)	1079	0.673
Total	1604	1.000

the diagnosis of FMF in 48.1% of these patients. The most common mutations were M694V, V726A, E148Q, and M680I(G/C). M694V was the most frequent mutation in patients with abdominal pain and arthritis in our study. Interestingly, M694V homozygosity and V726A heterozygosity were found more frequently in amyloidosis patients. The allele frequency of the M694V mutation was 0.0142, and a total of 195 (24.3%) patients had this mutation. M694V homozygotes accounted for 4.1% of the patients, 13.5% were M694V heterozygotes, and 17.6% were M694V compound heterozygotes. In previous studies, M694V was detected in 69-80% of FMF patients with arthritis and abdominal pain (Ince et al., 2002; Olgun et al., 2005). The M694V mutation is the most prevalent mutation in Arabs (Mattit et al., 2006; Jarjour, 2010), Lebanese, and Jordanians (Medlej-Hashim et al., 2000, 2005), Turks (Yalçinkaya et al., 2000; Güneşçar et al., 2005), Armenians (Cazeneuve et al., 1999), Iranian Azeri Turks (Esmaeili et al., 2008), and Jews (Dewalle, 1998; Touitou, 2001).

An association between homozygosity for the M694V mutation and arthritis in FMF patients has been reported by others (Brik et al., 1999; Kone et al., 2000; Olgun et al., 2005). The clinical presentation of arthritis in FMF has variable forms. The M694V mutation is most likely associated with arthritis, one of the severe complications of FMF. All arthritis patients in Central Anatolia should be evaluated for FMF to prevent the development of chronic arthritis, abdominal pain, and amyloidosis. Data on genotype-phenotype correlation in FMF generally agree with the presence of M694V homozygosity and its correlation with the most severe FMF phenotypes and amyloidosis (Dewalle et al., 1998; Cazeneuve et al., 1999; Kone et al., 2000; Olgun et al., 2005; Jarjour, 2010; Jarjour and Dodaki, 2011).

The most serious complication in FMF patients is chronic renal failure, which is highly associated with point mutations in the *MEFV* gene. The population in our study had a high rate of carriers, and V726A had the second-highest mutation frequency when our results were compared with those of other regions of Turkey and other Mediterranean groups (Shinar et al., 2000). Therefore, we expect that the M694V mutation, which causes the worst pathologic phenotype, must be the mutation with the least penetration in patients with FMF (Marek-Yagel et al., 2009; Jarjour and Dodaki, 2011). According to our study, chronic renal failure patients in the Central Anatolian population have a high rate of *MEFV* homozygous (M694V) and heterozygous (V726A) gene mutations.

In conclusion, *MEFV* gene mutations vary according to population characteristics such as familial history, parental consanguinity, migration, the presence of heterozygous carriers, and intrapopulation differentiation. Populations similar to the one in our study should be examined for mutation types and frequency to protect the health of future generations. Our study shows that the genetics of FMF are more complex than previously been appreciated and supports the mounting evidence that a single *MEFV* mutation may be associated with mild FMF symptoms. However, heterozygous patients presenting severe phenotypes should be further analyzed for less common second *MEFV* mutations using gene sequencing. In addition, our results have important implications for genetic counseling. We believe that this study will be helpful to clinicians by indicating the most frequently encountered FMF mutations in the Turkish population. Despite the current body of knowledge about FMF, prospective clinical studies with large numbers of patients across various ethnic groups will help to clarify this considerably debilitating disease.

The authors state that they have no conflict of interest.

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